

Plant Archives

Journal homepage: http://www.plantarchives.org

DOI Url: https://doi.org/10.51470/PLANTARCHIVES.2024.v24.no.2.226

EVALUATION OF FOUR TRICHOGRAMMATIDS AS BIOLOGICAL CONTROL AGENTS FOR *PECTINOPHORA GOSSYPIELLA* (SAUNDERS) IN INDIA

P.P. Dafare¹, V.J. Tambe¹, V. Chinna Babu Naik^{2*}, Prabhulinga T.³ and Jasti Sri Vishnu Murthy⁴

Department of Entomology, Dr. Panjabrao Deshmukh Krishi Vidyapeeth, Nagpur-440001, Maharashtra, India
 Division of crop protection, ICAR- Indian Institute of Rice Research, Hyderabad-500030, India
 Division of crop protection, ICAR- Central Institute for Cotton Research, Nagpur, Maharashtra 441108, India
 Department of Agricultural Entomology, College of Agriculture, Kerala Agricultural University, Vellanikkara, Thrissur-680656, Kerala, India.

*Corresponding author e-mail: chinnaenton@gmail.com (Date of Receiving-02-03-2024; Date of acceptance-23-05-2024)

ABSTRACT

Egg parasitoids (Trichogramma) play a crucial role in the pink bollworm *Pectinophora gossypiella* management. In this study, we assessed and compared the efficacy of four different Trichogramma species (*Trichogrammatoidea bactrae*, *Trichogramma chilonis*, *T. pretiosum*, and *T. achaeae*) on the eggs (0-, 1-, 2- and 3-day-old) of the pink bollworm, *P. gossypiella*, examining their parasitizing efficacy, emergence rates, and development periods under laboratory conditions. All four Trichogramma species parasitized *P. gossypiella* eggs at all stages of development, but they displayed varying preferences, showing a stronger inclination towards younger eggs. *T. bactrae* exhibited parasitization rates of 69%, 63%, 56%, and 65% on 0-, 1-, 2-, and 3-day-old eggs respectively within a 24-hour period, indicating its superior suitability and effectiveness across all egg ages. In contrast, *T. chilonis* showed the lowest parasitization and poorer host acceptance. Both *T. chilonis* and *T. pretiosum* displayed similar developmental periods when parasitizing eggs of all ages of *P. gossypiella*. However, *T. achaeae* and *T. bactrae* showed significantly slower development across all egg ages of *P. gossypiella*, although no significant differences were observed in adult emergence rates. These findings underscore the biological efficacy of *T. bactrae*, positioning it as a promising biocontrol agent against *P. gossypiella*. Consequently, it suggests its potential integration into bio-intensive Integrated Pest Management (IPM) strategies for cotton to mitigate the threat posed by *P. gossypiella*.

Key words: Biological control, Host age IPM, Pink bollworm, parasitization, Trichogramma,

Introduction

Cotton, a crucial commercial crop in India, occupies an extensive cultivation area of 130.49 lakh hectares, contributing substantially to global cotton production with an approximate share of 21% (ICAR-AICRP Cotton Annual Report, 2022-23). Cotton is a plant that nature seems to have designed specifically to attract insects (Rainwater, 1952). The indeterminate growth characteristics of the cotton crop offer food and shelter to a broad range of insects both directly and indirectly. Nearly 130 insect pest species plague Indian cotton, necessitating the management of around a dozen of these arthropods to optimize cotton yields, while globally, over

1300 herbivorous insects are identified in cotton systems (Hargreaves, 1948), though only a fraction are prevalent, and fewer still hold economic significance. Bollworms and sap sucking insects are important pests of cotton crop from the time it is sown until harvest. Bollworms, particularly the pink bollworm, are notorious for causing significant damage to cotton crop and reduce the economical yield (Ghosh, 2001). Several studies have shed light on the role of egg parasitoids, *Trichogramma* spp. in managing wide range of insect pests that affect cotton crops in different regions around the globe (Peter, 2022). Egg parasitoids (*Trichogramma*) are significant bio-agents in bio-intensive IPM strategy and successfully

1602 P.P. Dafare *et al*.

used against many larval pests including cotton bollworms (Cock, 1985; Ahmed et al., 1998; Malik, 2000; Charles et al., 2000 and Nadeem et al., 2009). Egg Parasitoid, *Trichogramma* is cosmopolitan in distribution and capable of parasitizing pink bollworm eggs (Ahmad et al., 2011). Various factors, such as adaptability to local environments (Pizzol et al., 2010, 2012; Zhang et al., 2014), effectiveness in dispersal (Ayvaz et al., 2008; Bueno et al., 2012), and preferences for host species or age (Zhu et al., 2014; Du et al., 2018), can significantly influence the field performance of Trichogramma parasitoids. It is imperative to examine and widely disseminate indigenous Trichogramma species for successful biological control programs, as they may possess unique characteristics tailored to specific host species and environmental conditions (Smith, 1996). Several studies have indicated that the efficiency of parasitization is impacted by the age of the host (Pak, 1986). Egg age emerges as a critical consideration when selecting appropriate Trichogramma species for pest biological control, as it affects the ethological and physiological dynamics of parasitoid-host interactions (Jarjees and Merritt, 2004; Pizzol et al., 2012); certain parasitoid species exhibit a preference for younger eggs.

In order to develop effective biological control programs, it is crucial to investigate the parasitization and suitability of Trichogramma parasitoids on *P. gossypiella* eggs at different stages of development. In this study, we assessed four Trichogramma species-*Trichogrammatoidea bactrae*, *T. chilonis*, *T. pretiosum*, and *T. achaeae*on *P. gossypiella* eggs under controlled laboratory conditions. The aim was to determine the parasitic capacity and suitability of these Trichogramma species for *P. gossypiella* eggs at various developmental stages, with the ultimate goal of identifying the most effective Trichogramma species for controlling this pest.

Materials and Methods

The experiment was carried out in the ICAR-CICR Insect Biocontrol facilities in Nagpur, India. The following *Trichogramma* species were utilized: *Trichogramma* achaeae (Nagaraja), *Trichogramma pretiosum* (Riley), *Trichogramma chilonis* (Ishii), and *Trichogrammatoidea bactrae* (Nagaraja). The factitious host employed for maintaining the culture was Corcyra cephalonica. Host culture (*Pectinophora gossypiella*) was maintained on artificial diet recommended by Naik *et al.*, 2017. Each species of *Trichogramma* was reared in glass vials as described by (Morison, 1970) at 27°C temperature and 65 per cent relative humidity was maintained.

Preliminary assays revealed that *P. gossypiella* eggs incubated for 4-5 days under laboratory conditions before hatching. Therefore, eggs aged 0, 1, 2, and 3 days were chosen for the study. Following the outlined procedure, mated females of each of the four Trichogramma species were initially introduced into separate replicated glass tubes, each containing a paper card with 100 pink bollworm eggs of a single age group (0-, 1-, 2-, or 3-dayold eggs). After allowing the female parasitoids to oviposit for 24 hours, they were removed. Eggs were then monitored for parasitization after being incubated for 8, 9, and 10 days, indicated by the darkening of the vitelline membrane. Since the eggs were not treated with UV, the number of host larvae hatching simultaneously was also recorded. Additionally, the eggs were inspected daily to observe the emergence of *Trichogramma* adults from the host eggs until no parasitoids emerged. Each Trichogramma species evaluated separately on various ages of pink bollworm eggs and was replicated four times. The parameters were calculated using the following equations.

$$\begin{aligned} \text{Per cent parasitization} &= \frac{\text{Number of Parasitized eggs}}{\text{Total number of host eggs}} \times \text{ 100} \\ \text{Per cent Adult emergence} &= \frac{\text{Number of eggs with emergence holes}}{\text{Total number of parasitized eggs}} \times \text{ 100} \end{aligned}$$

Developmental time (d) = represents the number of days from parasitization to wasp emergence

Data analysis

The per cent parasitized eggs and adult emergence and the development time were analyzed by a two-way ANOVA with parasitoid species (five levels) and host egg age (four levels) as factors at P<0.05.

Results and Discussion

Parasitization of *Trichogramma* species on *P. gossypiella* eggs at various ages

Trichogramma species and host egg age both significantly impacted the parasitization. Also, the interaction of egg age and Trichogramma spp on the proportion of parasitized eggs found significant ($F_{3,12}$ =6.004, P<0.001). Trichogramma species parasitization was significantly impacted by P. gossypiella egg age; as egg age grew, the % parasitization reduced.

T. chilonis had the highest parasitization 0-dayold eggs (72.25%) and parasitized fewer hosts with egg age increasing. *T. bactrae* had a similar parasitization on 0, 1, 2 and 3-day-old eggs. *T. pretiosum* and *T. achaeae* significantly preferred 1-day-old eggs to older eggs. There were significant differences in *Trichogramma* parasitization on various eggs ages with an exception of

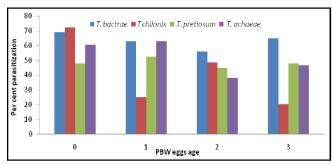


Fig. 1: Per cent of *P. gossypiella* eggs parasitized by the four *Trichogramma* species within 24 h at various ages.

0-day-old and 2-day-old eggs (0-day-old: $F_{3,12}=3.06$, P=0.06; 1-day-old: $F_{3,12}=15.42$, P<0.001; 2-day-old: $F_{3,12}=1.28$, P= 0.32 and 3-day-old: $F_{3,12}=8.37$, P<0.01).

Values in parentheses are arc sin transformed

T. bactrae had the highest parasitization on all host egg ages. More specifically, there were 69.00, 63.00, 56.00, and 65.00 host eggs parasitized in 24 h on 0-, 1-, 2-, and 3-day-old eggs, respectively. In general, *T. chilonis* parasitized the least host eggs at all egg ages compared to the other *Trichogramma* species (Fig. 1).

Development of the *Trichogramma* species on *P. gossypiella* eggs at various ages

The PBW egg age, the *Trichogramma* species, and the interaction of these factors significantly affected the parasitoid developmental time (Table 2). The developmental time of all *Trichogramma* species except for T. achaeae increased with the host egg age. In addition, when parasitoid females parasitized older P. gossypiella eggs, the development time of offspring development was shorter. T. chilonis and T. pretiosum had a similar developmental time on all ages of host eggs. The developmental time of different Trichogramma species was significantly different on various host egg ages (0-day-old eggs: $F_{3,12}$ =7.92, P<0.001; 1-day-old eggs: $F_{3,12}$ =5.88, P<0.001; 2-day-old: $F_{3,12}$ =8.54, P<0.001 and 3-day-oldeggs: $F_{3.12}=1.32$, P<0.001). At room temperature, the greatest developmental duration was recorded for T. achaeae (7.95 days), whereas Trichogrammatoidea bactrae, T. chilonis, and T. pretiosum took 7.25, 6.25, and 5.75 days, respectively. T. achaeae and T. pretiosum generally displayed the longest and shortest developmental times, respectively. There was highly significant difference found in the developmental stages of Trichogramma species on the pink bollworm eggs.

Adult Emergence of *Trichogramma* species on *P. gossypiella* eggs at various ages

PBW egg age and *Trichogramma* species had no significant effect on the Per cent of adult emergence.

Table 1: Results of the factorial ANOVA used to analyze (A) the numbers of *P. gossypiella* eggs parasitized (parasitization), (B) the Per cents of emergence of parasitoids from parasitized eggs, and (C) the developmental time of *Trichogramma* spp in *P. gossypiella* eggs. Main factors tested were the age of *P. gossypiella* eggs ("Host egg age" factor) and best performing *Trichogramma* spp for control of *P. gossypiella*

Parameters	Source of variation	Df	F	Pvalue		
	PS	3	6.99	< 0.001		
Per cent	HA	3	5.51	< 0.01		
Parasitization	PS*HA	9	6.00	< 0.001		
	Error	48				
Per cent adult emergence	PS	3	1.26	0.29		
	HA	3	0.53	0.66		
	PS*HA	9	0.94	0.50		
	Error	48				
Developmental period	PS	3	6.45	< 0.001		
	HA	3	1.95	< 0.001		
	PS*HA	9	8.05	< 0.001		
	Error	48				
*PS parasitoid species, HA host eggs						

However, the interaction between egg age and *Trichogramma* species had no significant effect (Table 2).

The parasitoid emergence rate increased in younger *P. gossypiella* eggs that were 0–1-day-old. The pattern was not exactly the same when *P. gossypiella* eggs were 2–3-day-old, the emergence rate decreased with the increased PBW eggs ages except for *T. pretiosum* and *T. achaeae* where the emergence rate increased; the highest emergence rate (91.51 %) was observed for *Trichogrammatoidea bactrae*. Despite different patterns between the four host egg ages tested, the two factors "Parasitoid" and "Host egg age" did not interact significantly. No significant differences were found on the emergence Per cent of *T. bactrae*, *T. chilonis*, *T. pretiosum* and *T. achaeae* from 0- to 3-day-old host eggs (Table 2).

The per cent of parasitized eggs, the number of adult emergences, the development period of female progeny, and the suitability of *Trichogramma* species are typically used to evaluate parasitization and suitability (Pak 1986; Miura and; Kobayashi *et al.*, 1998; Monje *et al.*, 1999; Takada *et al.*, 2000; Zhang *et al.*, 2014; Song *et al.*, 2015). Furthermore, the availability and acceptance of parasitoids as hosts are known to be influenced by the host's age (Vinson, 1976; Pak, 1986).

Four Trichogramma species were compared for

1604 P.P. Dafare *et al*.

Table 2: Comparisons of development time and per cent of emergence of the four *Trichogramma* species on differently aged *P.gossypiella* eggs.

Parameters	Species	PBW eggs age (day)				
		0	1	2	3	
Developmental period (day)	T. bactrae	8±0 ^b	9±0 ^d	6±0 ^b	6±0 ^b	
	T. chilonis	6±0a	7±0 ⁶	6±0 ^b	6±0 ⁶	
	T. pretiosum	6±0°	6±0a	6±0°	5±0°	
	T. achaeae	8±0 ^b	7±0°	8.75±0.25°	7±0°	
Adult Emergence (%)	T. bactrae	90.45 <u>+</u> 5.51(77.03)	95.90 <u>+</u> 1.36(79.85)	93.48 <u>+</u> 3.79(79.42)	86.19 <u>+</u> 13.80(77.99)	
	T. chilonis	86.20 <u>+</u> 7.22(71.74)	83.34+13.37(71.45)	75.23 <u>+</u> 9.80(63.91)	64.83 <u>+</u> 7.48(53.85)	
	T. pretiosum	84.38 <u>+</u> 6.54(67.77)	71.31 <u>+</u> 11.62(58.82)	91.88 <u>+</u> 2.89(75.64)	84.03 <u>+</u> 9.10(70.71)	
	T. achaeae	91.51 <u>+</u> 3.14(75.35)	76.26 <u>+</u> 15.64(63.53)	80.94 <u>+</u> 11.40(70.97)	86.97 <u>+</u> 4.68(71.61)	

Mean± SE are presented. Means in a column followed by the same lower case letter and means in a row followed by the same upper case letter do not differ significantly (P<0.05).

parasitization, adult emergence, and developing period on *P. gossypiella* eggs at varying ages. The results showed that *T. bactrae* had a higher rate of parasitization and a shorter developmental time than the other *Trichogramma* species. Moreover, *T. bactrae* had the greatest number of adults that emerged from parasitized *P. gossypiella* eggs at ages of 0-2 days. Out of all the examined *Trichogramma* species, *T. bactrae* had the most potential for suppressing *P. gossypiella*.

Three-day-old host eggs were found to be less parasitized than younger eggs, which is consistent with the theory that host egg age influences the parasitization rate of Trichogramma parasitoids (Pizzol, 2004). The outcomes of this study are consistent with those of other research (Brand et al., 1984; Calvin et al., 1997; Pak et al., 1986; Moreno et al., 2009). According to our research, all four of the native Trichogramma species are able to parasitize the youngest eggs, and they all accepted the P. gossypiella eggs at all ages. It appears that T. bactrae, T. chilonis, T. pretiosum, and T. achaeae were better suited to parasitize younger P. gossypiella eggs because they only demonstrated increased parasitization efficiency on 0 to 1-day-old eggs. Malik's (2001) findings, which indicated that Trichogrammatoidea bactrae could parasitize up to 95.71% of pink bollworm eggs which is in support to the parasitization rate (56-69%) in our study. The previous investigation revealed that T. bactrae had a strong parasitization potential and can parasitize 24 hours old eggs, or one day old eggs (87.66%), followed by 48 hours old eggs (73.50%), and the lowest Per cent was found in 72 hours old eggs (58.33%) (Asha et al., 2019). However, we found that T. bactrae apparently showed a relative low acceptance on 48 hours (56%) old eggs. T. achaeae exhibited good parasitic ability on all aged eggs of P. gossypiella, especial for 0 to 1-day-old eggs with a parasitization rate of 60.75 and 63 respectively suggesting that it was less adopted to old eggs. *T. chilonis* exhibited good parasitic ability on 0-day-old eggs with a parasitization rate of 72.25 per cent. Hou *et al.*, 2018 also found that *T. chilonis* exhibited similar preference for 0-2-day old *M. separata* eggs to 3-day-old eggs, suggesting that it was less adapted to old eggs.

Trichogramma parasitoids inject venom at the time of stinging, which eventually inhibits the developing host embryo from reaching adulthood, although they do not oviposit on host eggs older than 4-5 days (Benoit and Voegele, 1979). If older *P. gossypiella* eggs do not allow the host embryo to successfully develop, this could account for the reduced parasitization success in those eggs, leading to the collapse of the host egg.

According to Godfray (1994) and Vinson (1998), parasitoid females are known to evaluate the hosts they come into contact with. Their behavior is determined by cues they detect, particularly during ovipositor probing, or at the moment of stinging, though other mechanisms might also be at play (Outreman et al., 2001; Desneux et al., 2009a). It's possible that older eggs were rejected by Trichogramma females due to possible competition with host embryos that were already developed. Even though a parasitoid egg was deposited as a result of the probing, Asgari and Rivers (2011) report that the venom injected during the stinging likely caused the death of all probed host eggs, as has previously been documented in Trichogramma parasitoid species (Benoit and Voegele, 1979; Klomp et al., 1980). Further studies would have to be carried out to clarify this point.

For some parasitoids, the combination of host species and age may affect host selection (Vinson, 1976; Pak, 1986; Zhang *et al.*, 2014). According to Pizzol *et al.*, (2012), the *Trichogramma* species in the earlier experiments show varying acceptability for host eggs at varying ages. As a case study, *T. chilonis* favored eggs

that were intermediate in age (1-2 days old) (Pak, 1986; Miura and Kobayashi, 1998), while our findings showed that *T. chilonis* preferred *P. gossypiella* eggs that were 0 days old (Miura and Kobayashi, 1998). These variations imply that some parasitoids may pick their hosts based on the interaction between the host's age and species. Budhwant *et al.*, (2008) also reported that fresh 24-hour-old eggs (60.3%) had the best efficiency of *Trichogramma chilonis* Ishii against lepidopteran pests, followed by 48-hour-old eggs (49.5%) and 72-hour-old eggs (37.3%). It is consistent with the current study's findings that *T. chilonis* had the highest Per cent of parasitization at 0-day-old eggs (72.25%).

In this study, however, all *Trichogramma* species preferred to parasitize younger *P. gossypiella* eggs. This preference may lead to increase their offspring's survival or easily parasitize hosts. Pak (1986); and Honda and Luck (2000) reported that the first 75% of embryological development of the host egg was suitable for *Trichogramma* larvae development, although the pattern of vulnerability with host age varies among species. In our study, the four *Trichogramma* species had a lower Per cent of egg parasitized on 3-day-old *P. gossypiella* eggs. The possible mechanism underlying these phenomena is the differences in host quality associated with host egg age and the increasing defense capacity with the development of host egg (Mattiacci and Dicke, 1995).

Our results showed that Trichogramma species had high parasitization and good suitability on 0-1-day-old P. gossypiella eggs, and equal or more than one Trichogramma adults emerged from a single P. gossypiella egg. Our laboratory study demonstrated the P. gossypiella egg ages could affect the parasitization efficiency of the four Trichogramma species, providing valuable information for selecting suitable species to control P. gossypiella. In general, Trichogrammatoidea bactrae exhibits the highest parasitization capacity and suitability on all egg ages, and suites the best in controlling P. gossypiella. Practically, suitability is based on the ability of parasitoid to search for host eggs, the parasitoid quality, and the adaptability to pesticides and environment. Further research will be carried out under field conditions to measure the concrete indexes ensuring effective and practical biological control strategy.

Acknowledgement

The authors extend their heartfelt appreciation to Dr.Panjabrao Deshmukh Krishi Vidyapeeth and ICAR-Central Institute for Cotton Research for the invaluable support provided throughout their work. Their assistance, guidance, and resources have been instrumental in the successful completion of this project.

References

- Ahmad, N., Sarwar M., Muhammad R., Tofique M. and Wagan M.S. (2011). Conservation of bio-control agents in cotton, *Gossypium hirsutum* L. field by food supplements for insect pests management. *The Nucleus*, **48(3)**, 255-260.
- Ahmad, N., Ashraf M. and Fatima B. (1998). Potential of *Trichogramma chilonis* to parasitize eggs of pink, spotted andspiny bollworms of cotton. *Pak. J. Zool.*, **30(1)**, 39-40.
- Asgari, S. and Rivers D.B. (2011). Venom proteins from endoparasitoid wasps and their role in host-parasite Interactions. *Ann. Rev. Entomol.*, **56**, 313-335.
- Asha, S., Naik V.C.B., Neharkar P.S., Ughade J.D. and Sant S.S. (2019). Parasitizing potential of four Trichogramma species on the eggs of pink bollworm, *Pectinophora gossypiella* (Saunders). *J. Pharmacognosy Phytochem.*, **8(5)**, 857-859.
- Ayvaz, A., Albayrak S. and Karaborklu S. (2008). Gamma radiation sensitivity of the eggs, larvae and pupae of Indian meal moth *Plodiainterpunctella* (Hübner) (Lepidoptera: Pyralidae). *Pest Manag Sci.*, **64**, 505-512.
- Benoit, M. and Voegele J. (1979). Choix de l'ho^teetcomportementtrophique de *Trichogramma* evanescens Westw. (Hym., *Trichogramma*tidae) enfonction du de veloppementembryonnaired Ephestiakuehniella Zell. etd Ostrinianubilalis Hubner (Lep., Pyralidae). Entomophaga, **24**, 199-207.
- Brand, A.M., Dijken M.V., Kole M. and Lenteren J.V. (1984). Host-age and host-species selection of three strains of *Trichogramma evanescens* Westwood, an egg parasite of several lepidopteran species. *Meded Fac Land bouww Rijksuniv Gent.*, **49(3a)**, 839-847.
- Budhwant, N.P., Dadmal S.M., Nemade P.W. and Patil M.S. (2008). Efficacy of *Trichogramma chilonis* Ishii against Lepidopteran pests and age of host eggs. *Ann. Plant Sci.*, **16(1)**, 6-10.
- Calvin, D.D., Losey J.E., Knapp M.C. and Poston F.L. (1997). Oviposition and development of *Trichogramma pretiosum* (Hym., *Trichogrammatidae*) in three age classes of southwestern corn borer eggs. *Environ. Entomol.*, **26(2)**, 385-390
- Charles, P.C., Orr D.B., Van Duyn J.W. and Borchert D.M. (2000). *Trichogramma exiguum* (Hymenoptera: *Trichogrammatidae*) releases in North Carolina cotton: Evaluation of heliothine pest suppression. Department of Entomology, North Carolina State University, Raleigh, NC 2. 7695-7613.
- Cock, M.J.W. (1985). The use of parasitoids for augmentative biological control of pests in the people's republic of China. *Biocontrol News and Information.*, **6(3)**, 213-24.
- Desneux, N., Barta R.J., Hoelmer K.A., Hopper K.R. and Heimpel GE. (2009a). Multifaceted determinants of host specificity

1606 P.P. Dafare *et al*.

in an aphid parasitoid. Oecologia., 160, 387-398

- Desneux, N., Barta R.J., Delebecque C.J. and Heimpel G.E. (2009b). Transient host paralysis as a means of reducing self-super parasitization in koinobiontendoparasitoids. *J. Insect. Physiol.*, **55**, 321-327.
- Desneux, N., Blahnik R., Delebecque C.J. and Heimpel G.E. (2012). Host phylogeny and host specialization in parasitoids. *Ecol Lett.* **15**, 453-460.
- Du, W.M., Xu J., Hou Y.Y., Lin Y., Zang L.S., Yang X, Zhang J.J., Ruan C.C. and Desneux N. (2018). *Trichogramma* parasitoids can distinguish between fertilized or unfertilized host eggs. *J. Pest. Sci.*, **91**, 771-780
- Ghosh, S.K. (2001). GM crops: Rationally irresistible. *Curr. Sci.*, **81**(**6**), 655-660.
- Godfray, H.C.J. (1994). Parasitoids: behavioural and evolutionary ecology. Princeton University Press, Chichester.
- Hargreaves, H. (1948). List of the Recorded Cotton Insects of the World. London: Commonwealth Institute of Entomology.
- Honda, J.Y. and Luck. R.F. (2000). Age and suitability of Amorbiacuneana (Lepidoptera: Tortricidae) and Sabulodesaegrotata (Lepidoptera: Geometridae) eggs for *Trichogramma platneri* (Hymenoptera: *Trichogrammatidae*). *Biocontrol.*, **18**, 79-85.
- ICAR-AICRP (Cotton) Annual Report 2022-23. ICAR All India Coordinated Research Project on Cotton, Coimbatore. 641-003.
- Jarjees, E.A. and Merritt D.J. (2004). The effect of parasitization by *Trichogramma australicum* on *Helicoverpa armigera* host eggs and embryos. *J. Invertebr Pathol.*, **85**, 1-8.
- Klomp, H., Teerink B. and Wei C.M. (1980). Discrimination between parasitized and unparasitized hosts in the egg parasite *Trichogramma embryophagum* (Hym.: *Trichogrammatidae*): a matter of learning and forgetting. *Neth. J. Zool.*, **30(2)**, 254-277.
- Malik, F.M. (2000). Lifetable studies of *Trichogrammatoidea* bactrae (Hymenoptera: *Trichogrammat*idae) an effective biological agent of pink bollworm (*Pectinophora gossypiella*, Lepidoptera: Gelechiidae) of cotton (*Gossypium spp.*). Pakistan Journal of Biological Science, 3(12), 2106-2108.
- Malik, M.F. (2001). Field Release of *Trichogrammatoidea* bactrae, Hymenoptera: *Trichogrammatidae* an Effective Biological Agent of Pink Bollworm (Pectinophora gossypiella, Lepidoptera: Gelechiidae) of Cotton (Gossypium hirsutum L.). On Line Journal of Biological Sciences, 1(2), 56-57.
- Mattiacci, L. and Dicke M. (1995). The parasitoid Cotesiaglomerata (Hymenoptera: Braconidae) discriminates between first and fifth larval instars of its host Pierisbrassicae on the basis of contact cues from frass, silk, and herbivore-damaged leaf tissue. *J Insect Behav.*, **8**, 485-498.
- Miura, K. and Kobayashi M. (1998). Effects of host-egg age

- on the parasitization by *Trichogramma chilonis* Ishii (Hymenoptera: *Trichogrammatidae*), an egg parasitoid of the diamondback moth. *Appl. Entomol. Zool.*, **33**, 219-222
- Monje, J.C., Zebitz C.P.W. and Ohnesorge B. (1999). Host and host age preference of *Trichogramma galloi* and *T. pretiosum* (Hymenoptera: *Trichogrammatidae*) reared on different hosts. *J. Econ. Entomol.*, **92**, 97-103.
- Moreno, F., Perez-Moreno I. and Marco V. (2009). Effect of Lobesiabotrana (Lepidoptera: Tortricidae) egg age, density, and UV treatment on parasitization end development of T. cacoeciae (Hymenoptera: *Trichogrammatidae*). Environ. Entomol., **38**, 1513-1520.
- Morison, R.K. (1970). A simple cage for maintaining parasites. *Ann. Entomol. Soc. Am.* **63**, 625.
- Nadeem, S., Ashfaq M., Hamed M., Ahmed S. and Kashif M. (2009). Comparative rearing of *Trichogramma chilonis* (Ishii) (Hymenoptera: *Trichogrammatidae*) at different temperature conditions. *Pakistan Entomologist* **31(1)**, 33-36.
- Naik, V.C.B., Kranthi S., Kumbhare S. and Nagrare V.S. (2017).
 A manual on pink bollworm resistance monitoring and management. ICAR-CICR, Technical Bulletin.
- Outreman, Y., Le Ralec A., Plantegenest M., Chaubet B. and Pierre J.S. (2001). Super parasitization limitation in an aphid parasitoid: cornicle secretion avoidance and host discrimination ability. *J. Insect. Physiol.*, **47**, 339-348.
- Pak, G.A. (1986). Behavioural variations among strains of *Trichogramma* spp: a review of the literature on hostage selection. *J. Appl. Entomol.*, **101**, 55-64.
- Peter, A. Edde, (2022). 4 Arthropod pests of cotton (*Gossypium hirsutum* L.), Editor (s): Peter A. Edde, Field Crop Arthropod Pests of Economic Importance, Academic Press, 2022, Pages 208-274
- Pizzol, J., Pintureau B., Khoualdia O. and Desneux N. (2010). Temperature dependent differences in biological traits between two strains of *Trichogramma cacoeciae* (Hymenoptera: *Trichogrammatidae*). *J. Pest. Sci.*, **83**, 447-452.
- Pizzol, J. (2004). Etudes bioe´cologiques de *Trichogramma* cacoeciae Marchal, parasitoý¨deoophage de l'eude´mis de la vigne, envue de son utilisationenluttebiologique. Diploˆmed'Inge´nieurDiploˆme´ par l'Etat, option Agriculture ENSAM, Montpellier.
- Pizzol, J., Desneux N., Wajnberg E. and Thiery D. (2012). Parasitoid and host egg ages have independent impact on various biological traits in a *Trichogramma* species. *J. Pest Sci.* **85**, 489-496.
- Rainwater, C.F. (1952). Progress in research on cotton insects. In Insects, The Yearbook of Agriculture, 1952, 497-500. Washington, DC: US Department of Agriculture, US Government Printing Office.
- Smith, S.M. (1996). Biological control with *Trichogramma*: advances, success, and potential of their use. *Ann Rev Entomol.*, **41**, 375-406.

- Song, L.W., Wen X.Y., Zhang L.S., Ruan C.C., Shi S.S., Shao X.W. and Zhang F. (2015). Parasitization and suitability of different egg ages of the *Leguminivora glycinivorella* (Lepidoptera: Tortricidae) for three indigenous *Trichogramma* species. *J. Econ. Entomol.*, 108, 933-939.
- Takada Y., Kawamura S. and Tanaka T. (2000). Biological characteristics. Growth and development of the egg parasitoid *Trichogramma dendrolimi* (Hymenoptera: *Trichogrammatidae*) on the cabbage armyworm Mamestrabrassicae (*Lepidoptera: Noctuidae*). *Appl. Entomol. Zool.*, **35**, 369-379.
- Vinson, S.B. (1976). Host selection by insect parasitoids. *Ann. Rev. Entomol.*, **21**, 109-133.

- Vinson, S.B. (1998). The general host selection behavior of parasitoid hymenoptera and a comparison of initial strategies utilized by larvaphagous and oophagous species. *Biol. Control*, **11**, 79-96.
- Zhang, J.J., Ren B.Z., Yuan X.H., Zang L.S., Ruan C.C., Sun G.Z. and Shao X.W. (2014). Effects of host-egg ages on host selection and suitability of four Chinese *Trichogramma* species, egg parasitoids of the rice striped stem borer. *Chilosuppressalis Bio. Control.*, **59**, 159-166.
- Zhu, P., Zhang Y.F., Song Q.T., Zhang F. and Li Y.X. (2014). The suitability of Ostriniafurnacalis (Lepidoptera: Crambidae) eggs for *Trichogramma dendrolimi* (Hymenoptera: *Trichogrammatidae*) can be changed by T. ostriniae. *Appl. Entomol. Zool.*, **49**, 265-272.